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A NEW PENTACYCLIC TRITERPENE LACTONE FROM *DILLENIA INDICA**

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Key Word Index—*Dillenia indica*; Dilleniaceae; myricetin; sitosterol; betulinic acid; betulinaldehyde; betulin; lupeol; a new hydroxylactone— 3β -hydroxy-lupane-13 β , 28-lactone.

Plant. *Dillenia indica* Linn is an evergreen tree native to India. *Uses.* Its bark and leaves are astrigent [1]. *Previous work.* On trunk bark [2], on leaves [3].

Present work. Air-dried powdered stem-bark was successively extracted with light petrol, CHCl_3 and MeOH. Light petrol extract was dried and separated into neutral and acidic fractions by the usual method. The neutral fraction on repeated chromatography on SiO_2 column afforded betulinaldehyde, m.p. 199–200° $[\alpha]_D^{26} + 28^\circ$,[†] betulin, m.p. 254–255° $[\alpha]_D^{27} + 22^\circ$, Lupeol, m.p. 211–212° $[\alpha]_D^{29} + 27^\circ$, sitosterol, m.p. 137° $[\alpha]_D^{28} - 36^\circ$ and the acidic fraction yielded only betulinic acid, m.p. 305–306° $[\alpha]_D^{27} + 6.35^\circ$. These compounds were identified by comparison with authentic samples (m.m.p., co-TLC, superimposable IR). The isolation of betulinic acid, betulinaldehyde, betulin and lupeol showed a remarkable biogenetic sequence rarely encountered in a plant source. The CHCl_3 extract yielded only betulinic acid and betulin.

The dried MeOH extract was re-extracted with CHCl_3 to remove nonglycosides and the mixture of glycosides, which could not be separated on SiO_2 , was hydrolysed with 6% methanolic HCl. The only sugar identified was D-glucose (PC). The aglycone fraction was separated into acidic and neutral parts. The acidic part yielded only betulinic acid, and the neutral part, on column chromatography on SiO_2 , afforded sitosterol, betulin,

hydroxylactone B of betulinic acid [4] along with the flavonol myricetin, m.p. 358–360°, acetate, m.p. 214–216° and identified by comparison with an authentic sample (m.m.p., Co-TLC, superimposable IR, UV). Hydroxylactone B was most probably an artefact formed by influence of acid on betulinic acid.

Besides this series of known compounds $\text{C}_6\text{H}_6:\text{CHCl}_3$ (4:1) eluate from the chromatogram yielded a triterpene (+ve Liebermann–Burchard test –ve, $\text{C}(\text{NO}_3)_4$ m.p. 325° (d), $[\alpha]_D^{27} + 63.4^\circ$, $\text{C}_{30}\text{H}_{48}\text{O}_3$ (Found: C, 78.45; H, 10.42%; Calc. for $\text{C}_{30}\text{H}_{48}\text{O}_3$, C, 78.89; H, 10.59%) MW 456 by MS. Acetate m.p. 319–320° $[\alpha]_D^{27} + 82.1^\circ$. IR, 1754 cm^{-1} (5-membered lactone) and 3350 cm^{-1} (hydroxyl), suggestive of a hydroxylactone. This lactone was different from hydroxylactone A and B by comparison with authentic samples [4].

On oxidation with Jones's reagent it gave a keto compound, m.p. 328–330° $[\alpha]_D^{27} + 70^\circ$; IR, 1689 and 1754 cm^{-1} . The ketone gave a positive Zimmerman's test and was reduced back to the original compound with NaBH_4 which proved the β conformation of OH group at C-3. Studies of its MS pattern [5] (prominent peak for $M - 43$ unit for isopropyl) NMR spectra [6] and also on biogenetic ground, indicated the compound to be in the lupane series. It was not identical with dihydrothuberogenin [7] or the dihydrolactone produced by mercuric acetate oxidation of acetyl betulinic acid [8].

The compound was reduced by LiAlH_4 to give a triol m.p. 280–281° $[\alpha]_D^{27} + 34^\circ$, (Found: C,

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[†] Unless otherwise stated, all $[\alpha]_D$ values are in CHCl_3 .

78.31; H, 11.35%, Calc. for $C_{30}H_{52}O_3$, C, 78.20; H, 11.38%). IR spectra showed no CO. Acetylation (at room temperature and at 130°) gave a diacetate (without any trace of triacetate) m.p. 230–231° $[\alpha]_D^{27} + 32^\circ$ (found: C, 74.89; H, 10.42; acetyl, 15.59, calc. for $C_{34}H_{56}O_5$, C, 74.95; H, 10.36; acetyl, 15.72%). Stability of this diacetate towards CrO_3 proved that the OH-group must be tertiary one i.e. at C – 13. The triol diacetate was dehydrated with BF_3 in dry benzene [9], purified on a $AgNO_3$ impregnated silica gel column, to give a major fraction which on crystallization ($CHCl_3$ –MeOH) yielded a pure compound m.p. 295–296° $[\alpha]_D^{27} + 70.2^\circ$. Although the compound gave yellow colour with $C(NO_3)_4$, its IR spectra showed no absorption for a trisubstituted double bond. UV210 ($\epsilon = 5650$) 215 (4520) and 220 nm (3650), were indicative of a tetrasubstituted double bond [10]. On oxidation with CrO_3 in HOAc [11] a conjugated ketone was obtained, m.p. 290° $[\alpha]_D^{27} + 65^\circ$. IR 1690 cm^{-1} , and UV, at 242 nm ($\epsilon = 13520$) without any absorption for vinylic proton in NMR. Thus the position of double bond introduced by dehydration of triol-diacetate is most likely to be at C 13(18). In view of the evidence outlined above the structure of the new hy-

droxylactone is suggested to be 3 β -hydroxy-lupane-13 β -28-lactone (1).

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ASEBOTIN AND ITS AGLUCONE FROM THREE SPECIES OF RHODODENDRON

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Key Word Index—*Rhododendron canescens*, *R. nudiflorum*, *R. roseum* and *R. calendulaceum*; Ericaceae; dihydrochalcones; asebotin; 2',6',4-trihydroxy-4-methoxydihydrochalcone.

In the course of a biochemical systematic investigation of *Rhododendron* (Ericaceae), we isolated two dihydrochalcones, one of which was not previously reported from Nature. In a recent review,

Bohm [1] noted that there are only 13 naturally occurring dihydrochalcones including phloridzin (2',4',6',4-tetrahydroxydihydrochalcone 2'-O-glucoside) and asebotin (the 4'-O-methyl ether of